

# Endocrine changes after experimental showjumping

Adriana Ferlazzo\*, Pietro Medica, Cristina Cravana and Esterina Fazio

Department of Morphology, Biochemistry, Physiology and Animal Production, Unit of Veterinary Physiology, Faculty of Veterinary Medicine, University of Messina, 98168 Messina, Italy

\* Corresponding author: ferlazzo@unime.it

Submitted 25 May 2009; Accepted 14 September 2009

Research Paper

## Abstract

The study was designed in order to gain a better understanding of whether the lack of competition stress and/or sampling time had an influence on circulating  $\beta$ -endorphin, adrenocorticotrophin (ACTH) and cortisol modifications after experimental showjumping sessions and to study the effects of fence height on hormone changes. Hormone levels were recorded before exercise in basal conditions and after warm-up, then 5 and 30 min post-exercise. Using a randomized crossover study design, six horses were studied during three experimental showjumping sessions over fences of different heights: 1.00 m (session 1), 1.10 m (session 2) and 1.20 m (session 3). The showjumping exercise did not modify plasma  $\beta$ -endorphin and serum ACTH concentrations after session 1, and tended only to maintain higher values than basal after both session 2 and session 3. The interaction fence height/time was not statistically significant for either  $\beta$ -endorphin or ACTH changes. Sampling time significantly affected both  $\beta$ -endorphin ( $F = 2.88$ ;  $P < 0.04$ ) and ACTH ( $F = 3.84$ ;  $P < 0.01$ ) changes. Serum cortisol levels were always higher than basal 5 min post-exercise, with levels falling at 30 min. The interaction fence height/time was not statistically significant, while sampling time significantly affected the results ( $F = 7.96$ ;  $P < 0.0002$ ). This study demonstrated no significant effects of fence height on  $\beta$ -endorphin, ACTH and cortisol changes. The sampling times adopted affected post-exercise changes in plasma  $\beta$ -endorphin, ACTH and cortisol and could have masked the effects of fence height on hormone modifications.

**Keywords:** horse; showjumping;  $\beta$ -endorphin; ACTH; cortisol

## Introduction

In order to maintain homeostasis under exercise stress, the hypothalamus-hypophysis-adrenocortical (HPA) axis is activated together with the autonomic nervous system. The secretion of adrenocorticotrophin (ACTH) and cortisol is modulated by the intensity, duration and type of exercise, as extensively reported<sup>1–8</sup>.

Endorphins are endogenous, opiate-like peptides of pituitary origin that modulate pain perception and they also play a role in the response to physical and psychological stress. In man, the heightened activation of the HPA axis and increased  $\beta$ -endorphin modulate the time to fatigue and are associated with performance impairment<sup>9</sup>.

In horses, plasma  $\beta$ -endorphin, ACTH and cortisol are considered to be reliable physiological markers for welfare<sup>10–12</sup> and performance/fitness evaluation<sup>13</sup>. The association of plasma  $\beta$ -endorphin concentrations

with the severity of exercise stress has also been studied<sup>14,15</sup>.

It has been reported that increases in plasma cortisol induced by ACTH administration could be used to predict the exercise-induced stress response<sup>16</sup>. It is widely known that physical exercise generally induces a clear-cut increase in circulating cortisol levels, even from activities requiring exercise of mild intensity and short duration, such as showjumping<sup>3,5,17</sup>.

In Thoroughbreds, exercising on a treadmill, the response of cortisol to an incremental exercise and two constant relative workload exercises (105 and 80%  $VO_{2max}$ ) appeared to be correlated with the duration of exercise but were not correlated to plasma ACTH<sup>18</sup>. However, cortisol secretion from the adrenal cortex in the horse is generally promoted by ACTH administration and/or endogenous release<sup>2,19</sup>.

It is well known that the cortisol response usually depends on the state of individual fitness and

training<sup>17,20-22</sup>. Moreover, training may produce a more rapid decline in cortisol levels after exercise towards resting values<sup>2</sup>, while overtraining has been associated with a reduction in the cortisol response to exercise<sup>23,24</sup>. It has been reported that cortisol and  $\beta$ -endorphin modifications usually occur in phase in response to exercise<sup>25</sup>.

Age as well as training has been reported to affect  $\beta$ -endorphin basal levels and the hormonal response to acute exercise<sup>13,26,27</sup>. Falling ACTH and  $\beta$ -endorphin values were recorded after training<sup>13</sup>. After overtraining<sup>24</sup>, the mean  $\beta$ -endorphin concentrations in resting horses were not significantly different.

Exercise produces significantly increased concentrations in  $\beta$ -endorphin<sup>13,28</sup>. Treadmill exercises at submaximal levels of exercise produced significant increased concentrations of  $\beta$ -endorphin in horse plasma, while ACTH levels did not change<sup>13</sup>. In contrast, acute vigorous physical exercise promptly heightened both pituitary venous blood ACTH as well as vasopressin, the main ACTH secretagogue<sup>29</sup>. Moreover, increased plasma ACTH concentrations have been recorded in response to exhaustive incremental treadmill exercise<sup>30</sup>. The ACTH response found was more sensitive to the intensity of exercise and showed a significant correlation with blood lactate concentrations<sup>18</sup> and with plasma vasopressin concentrations during incremental exercise tests<sup>31</sup>.

The effect of exercise duration on  $\beta$ -endorphin modifications was found to be highly dependent on the type of exercise performed<sup>32</sup>. During incremental exercise tests, plasma  $\beta$ -endorphin concentrations were positively correlated with both exercise speed<sup>13,33</sup> and intensity<sup>14</sup>. A critical threshold intensity of  $\leq 60\%$   $\text{VO}_{2\text{max}}$  for significant increases in  $\beta$ -endorphin concentrations in horses exercising on a treadmill for 5 min or longer has been reported<sup>15</sup>.

Showjumping is quite an intense sport despite the relatively slow average speed. This is due in part to the intense effort required to jump fences every 5 s. Despite this, the cortisol increase in showjumpers was found to be lower than in racehorses<sup>3,17</sup>.

Power, speed and technical jumping skill influence how successfully horses perform<sup>34</sup>. Smaller increases in plasma cortisol were found in experienced jumpers than in inexperienced jumpers, suggesting that horses become conditioned to the psychological stress of the show environment<sup>35</sup>. In competing horses, exercise stress is determined by a mix of different factors depending on the characteristics of the exercise and on individual fitness level and perception of stress conditions. In fact, resting cortisol levels of sport horses appear to be affected by emotional stress prior to competition<sup>5</sup>. Emotional stress in the form of novelty stimuli during exercise tests was also shown to increase plasma ACTH<sup>31</sup>.

Moreover, the circuit design specific to showjumping incorporates a number of additional exercise features to cope with, represented by the variety, number and height of fences. It requires that horses are able to continuously adapt their cardiovascular, respiratory and musculoskeletal systems in order to modulate the intensity of speed as well as to demonstrate their technical jumping skill.

Previous investigations on jumpers suggested that jumping ability as well as other competitive factors (e.g. size of fences) could influence serum cortisol changes, while circulating ACTH and  $\beta$ -endorphin were not so affected<sup>36,37</sup>.

Based on the above, in order to gain a better understanding of whether the lack of competition stress and/or sampling time had an influence on hormone modifications after showjumping and to study whether there are effects of fence height on hormone changes after exercise, this research studied the response of circulating  $\beta$ -endorphin, ACTH and cortisol concentrations in horses after three experimental showjumping sessions by evaluating the effects on hormone levels caused by the increasing fence heights used in the exercises.

## Materials and methods

### *Horses and experimental design*

Six trained sport jumper horses (three Sella Italiano, one Belgian Warmblood, one Dutch Warmblood and one Selle Français), three geldings, two mares and one stallion with a mean age of  $10.3 \pm 3.4$  years (range 6–13) and weight  $580 \pm 50$  kg were used. All horses were housed individually in  $4 \times 4$  stables at the Equine Research Facility where the test was carried out.

Each horse was fed a daily ration of 8 kg of alfalfa and grass hay and 4 kg of a commercially available hay cube ration (split into two feeds). Water and a salt/mineral block were provided in the boxes *ad libitum*.

The horses used in the study were able to cope with fences up to 1.30 m high and they all had previous experience of showjumping competition at the same level. The horses were under regular training and were considered by the referring veterinarian to be clinically healthy before entering the study. The horses and rider usually trained together approximately 1–2 h per day, performing low-level jumping exercises approximately twice a week, and were familiar with the jumping exercises required.

The experimental study was designed to limit the emotional stress arising from a competition environment and to exclude the influence of horses having differing degrees of competition experience.

Using a randomized crossover study design, the horses performed the three experimental jumping sessions over fences of different heights: 1.00 m (session 1), 1.10 m (session 2) and 1.20 m (session 3) over three consecutive days. The jumping sessions were performed after a warm-up. The warm-up consisted of a 10 min walk, trot and quiet canter. The horses jumped two low fences before tests commenced. After the warm-up, the horses walked for 10 min until the heart rate decreased to  $< 50$  beats  $\text{min}^{-1}$ .

The horses performed the three different sessions with the same rider. All the tests were performed in an outdoor arena before the morning feed and the mean ambient temperature was  $12^{\circ}\text{C}$  (range  $10$ – $15^{\circ}\text{C}$ ). The horses cantered an 800 m course and the circuit design differed exclusively on the basis of fence height. Five upright and five cross-pole fences were utilized in each session. The mean speed during the tests was  $5.5 \pm 0.3$   $\text{m s}^{-1}$ .

Heart rate was continuously monitored with a Polar Sport Tester (Polar Electro Europe BV, Fleurier, Switzerland).

Blood samples before and after the tests were taken between 07.00 and 09.00 hours, in order to minimize the effects of diurnal rhythms.

Blood samples (10 ml) were collected by jugular venipuncture using evacuated tubes (Venoject, Terumo<sup>®</sup>, Leuven, Belgium), while horses were in their stalls at rest daily at 07.00 hours, before both feeding and exercise. The same operator performed the procedure under quiet conditions and the horses were not restrained during it. All the other samples were collected after warm-up, then 5 and 30 min after the end of showjumping sessions.

In order to analyse  $\beta$ -endorphin concentrations, after collection an aliquot of the blood samples (5 ml) was transferred into polypropylene tubes containing EDTA ( $1 \text{ mg ml}^{-1}$  of blood) and aprotinin ( $500 \text{ KIU ml}^{-1}$  blood, ICN Biomedicals Inc., Aurora, OH, USA) and kept at  $4^{\circ}\text{C}$ . Plasma samples were harvested after centrifugation at 3000 for 15 min at  $4^{\circ}\text{C}$  and stored at  $-80^{\circ}\text{C}$  until the day of analysis. Peptides were extracted from plasma samples with 1% trifluoroacetic acid (TFA, HPLC grade) and elution with 60% acetonitrile (HPLC grade) in 1% TFA.

Plasma  $\beta$ -endorphin concentrations were measured in duplicate utilizing a commercial RIA kit (Peninsula Lab. Inc., Belmont, CA, USA) for human  $\beta$ -endorphin, which has a 100% cross-reactivity with equine  $\beta$ -endorphin<sup>14,15,38</sup>. The hormone assay utilized has a range for the amount of  $\beta$ -endorphin detected of  $1$ – $128 \text{ pg } 100 \mu\text{l}^{-1}$  ( $3$ – $371 \text{ pmol l}^{-1}$ ). The respective inter- and intra-assay CVs were 7 and 15%.

For the analysis of serum ACTH and cortisol concentrations, the blood samples (5 ml) were centrifuged at

$3000 \times g$  for 15 min at  $4^{\circ}\text{C}$  after collection. Serum samples were then harvested and stored at  $-20^{\circ}\text{C}$  and assayed for ACTH and cortisol.

Serum ACTH concentrations were analysed in duplicate using a commercially available RIA kit (ELSA-ACTH, CIS-BioInternational, Gif-sur-Yvette, France) suitable for equine use<sup>36</sup>. The hormone assay utilized has a range of the amount of ACTH detected of  $0$ – $2000 \text{ pg ml}^{-1}$  ( $0$ – $440 \text{ pmol l}^{-1}$ ). The inter- and intra-assay CVs were 6 and 15%, respectively.

Serum cortisol concentrations were analysed in duplicate using a commercially available immunoenzymatic kit (Roche Diagnostics GmbH, Mannheim, Germany). The hormone assay utilized has a range for the amount of cortisol detected of  $0$ – $1380 \text{ nmol l}^{-1}$ . The inter- and intra-assay CVs were 4.6 and 6.9%, respectively.

All methods and procedures used in this experiment were reviewed and approved by the Messina University Institutional Board for the Care and Use of Animals.

### Statistics

Data are presented as mean  $\pm$  SD.

Results were analysed using a statistical analysis software programme (SAS/STAT, Version 8.0; SAS Institute Inc., Cary, NC, USA). A one-way ANOVA for repeated measures (RM-ANOVA) was applied to test for any differences in the basal values on the three experimental days and to test for any differences in values after warm-up on the three experimental days. A two-way ANOVA for repeated measures (two-way RM-ANOVA) was applied to test for the effects of different fence heights and sampling times. When the  $F$  statistic was significant, the differences between individual means over time were then assessed using a *post hoc* multiple comparison test (Bonferroni).

The percentage differences ( $\Delta\%$ ) of changes in hormone concentrations were also calculated by comparing the different post-exercise times with basal levels.

Correlations between cortisol and  $\beta$ -endorphin, cortisol and ACTH and  $\beta$ -endorphin and ACTH were evaluated using linear regression analysis.

Statistical significance was set at  $P < 0.05$ .

## Results

### Heart rate

The horses had a mean heart rate of  $93.7 \pm 6.3$  beats  $\text{min}^{-1}$  after warm-up. The respective mean peak heart rates during the sessions were:  $175 \pm 15$  in session 1,  $182 \pm 10$  in session 2 and  $185 \pm 18$  beats  $\text{min}^{-1}$  in session 3. During the recovery, heart rate decreased to a mean of  $70.5 \pm 9.5$  beats  $\text{min}^{-1}$  after 5 min and was  $< 50$  beats  $\text{min}^{-1}$  after 30 min.

### $\beta$ -Endorphin

The effects of experimental jumping exercise sessions on  $\beta$ -endorphin modifications are presented in Fig. 1.

Basal and post-exercise  $\beta$ -endorphin values showed a wide range of individual variability.

RM-ANOVA showed that the mean basal  $\beta$ -endorphin levels of horses did not show any statistically significant changes over the 3 days of activity. After warm-up, the only significant increases in comparison with basal levels were recorded for session 2 ( $P < 0.01$ ).

In session 1, the jumping exercise did not modify plasma  $\beta$ -endorphin concentrations. In session 2, higher but not statistically significant post-exercise values than basal were recorded at both 5 and 30 min, although they were lower than warm-up levels. In session 3, no statistically significant higher post-exercise values than basal and warm-up were recorded at 5 min, and levels declined at 30 min.

Two-way RM-ANOVA showed the interaction fence height/time not to be significant. However, sampling time did significantly affect  $\beta$ -endorphin changes ( $F = 2.88$ ;  $P < 0.04$ ).

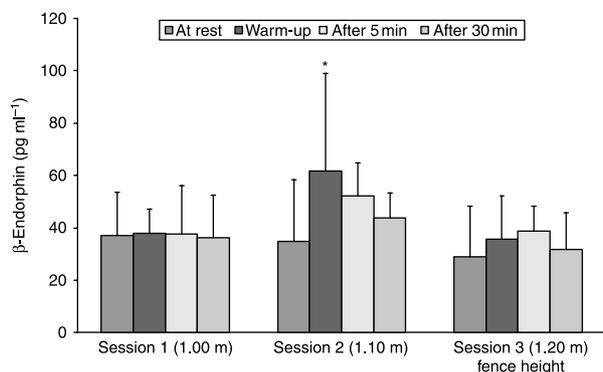
### Adrenocorticotrophin

The effects of experimental jumping exercise sessions on ACTH modifications are presented in Fig. 2.

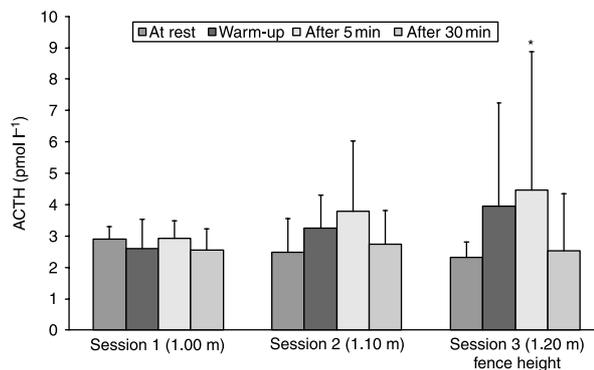
RM-ANOVA showed that the mean basal ACTH levels of horses did not statistically significantly decrease over the 3 days of activity.

At warm-up, non-statistically significant increases in comparison with basal levels were recorded, but only in session 2 and session 3, with values showing wide-ranging individual variability in session 3.

The jumping exercise did not modify serum ACTH concentrations after session 1. After session 2, ACTH showed higher basal levels 5 min post-exercise, and values declined after 30 min. After session 3, ACTH showed statistically significant higher values than basal 5 min post-exercise ( $P < 0.05$ ) and declining



**Fig. 1** Plasma  $\beta$ -endorphin concentrations (mean  $\pm$  SD) of horses ( $n = 6$ ) before and after sessions with various fence heights. Asterisk indicates significant ( $P < 0.01$ ) differences in average hormone *versus* basal in each session



**Fig. 2** Serum ACTH concentrations (mean  $\pm$  SD) of horses ( $n = 6$ ) before and after sessions with various fence heights. Asterisk indicates significant ( $P < 0.05$ ) differences in average hormone *versus* basal in each session

levels at 30 min. In both session 2 and session 3, 5 min post-exercise ACTH values showed wide-ranging individual variability.

The interaction fence height/time was not statistically significant. However, sampling time significantly affected ACTH changes ( $F = 3.84$ ;  $P < 0.01$ ).

The increases in the two hypophyseal hormones,  $\beta$ -endorphin and ACTH, recorded at 5 min post-exercise were equivalent (+51.0%) when horses jumped 1.10 m fences (session 2), while they were lower for  $\beta$ -endorphin (+33.7%) than for ACTH (+93.1%) when the horses performed a showjumping session over 1.20 m fences (session 3). No correlation between  $\beta$ -endorphin and ACTH was detected when the values for each test were analysed.

### Cortisol

The effects of experimental jumping exercise sessions on cortisol modifications are presented in Fig. 3.

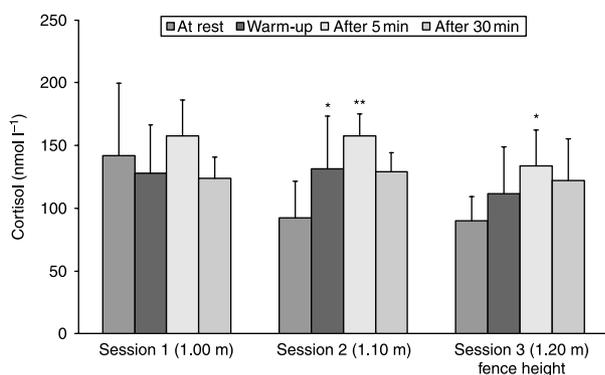
RM-ANOVA showed the mean basal levels of cortisol of horses did not statistically significantly decrease over the 3 days of activity.

At warm-up, significant increases in comparison with basal levels were recorded in session 2 ( $P < 0.05$ ).

Cortisol levels showed higher values than basal 5 min post-exercise over the course of the three showjumping sessions. The values at 30 min showed a drop in levels, although they still tended to be higher than basal after session 2 and session 3. With respect to basal conditions, significant differences were recorded at 5 min post-exercise in session 2 ( $P < 0.001$ ) and in session 3 ( $P < 0.05$ ).

The interaction fence height/time was not found to be statistically significant. However, the effect of sampling time was highly statistically significant ( $F = 7.96$ ;  $P < 0.0002$ ).

Statistically positive significant correlations emerged between cortisol and  $\beta$ -endorphin changes in session 1 ( $r = 0.47$ ;  $P < 0.029$ ) and between cortisol and ACTH changes in session 3 ( $r = 0.52$ ;  $P < 0.001$ ).



**Fig. 3** Serum cortisol concentrations (mean  $\pm$  SD) of horses ( $n=6$ ) before and after sessions with various fence heights. Asterisks indicate significant ( $*P < 0.05$ ;  $**P < 0.001$ ) differences in average hormone *versus* basal in each session

## Discussion

The results of this study demonstrated that experimental showjumping sessions induced increases in cortisol levels 5 min post-exercise, which were statistically significant only after performances over fence heights of 1.10–1.20 m (session 2 and session 3), and declining levels at 30 min, although values remained still higher than basal after session 2 and session 3, probably as an effect of the higher fences. No significant ACTH and  $\beta$ -endorphin modifications were recorded at 5 min post-exercise, with increases being found, once again, exclusively after showjumping sessions over 1.10–1.20 m fences (session 2 and session 3). At 30 min post-exercise, both ACTH and  $\beta$ -endorphin values tended to decline.

The degree of post-exercise increases was not equivalent for the different variables studied, and therefore the influence of haemoconcentration would not account for the  $\beta$ -endorphin, ACTH and cortisol changes recorded post-exercise.

The general increase in circulating cortisol levels recorded in horses after the three experimental sessions was consistent with the results usually reported as an effect of different types of exercise<sup>1,4–8</sup>, as well as after showjumping<sup>3,5,35,36,39</sup>. In the various reports, the entity of post-exercise increases recorded generally differed due to the timing of blood sampling or the differing levels of horse competition experience.

Therefore, the post-exercise cortisol increases recorded in this study represent mainly the physiological challenge response to the physical stress of exercise. In response to exercise, the cortisol increase affects substrate mobilization and cardiopulmonary function and it also prepares the system for the post-exercise recovery period<sup>40</sup>. Moreover, it is well known that jumping as an exercise normally involves a substantial physical stress component<sup>34</sup>.

In this study, the cortisol increase evaluated 5 min post-exercise in the horses studied did not show any

clear-cut differences compared with competitive showjumping sessions previously studied<sup>8</sup>. Nevertheless, on evaluating the post-exercise cortisol changes in horses with different levels of competition experience<sup>36</sup>, it was shown that the less experienced horses had higher post-exercise cortisol increases after competition, even for jumping over 1.00 m fences, than those recorded in the experimental session over the same fence height in this present procedure, probably due to the previous competition experience of horses studied. Cortisol and ACTH responses to jumping in competition have previously been shown<sup>35,36,39</sup> to be much lower with more experienced horses.

Moreover, in this experimental study, the mix of individual factors (experience, environmental stress and effect of the rider) that normally contribute to affect the involvement of HPA axis during competition were essentially kept under control, and therefore the psychological stress that is usually elicited by exercise performed in competitive conditions was probably slight.

The interaction fence height/time was not statistically significant for cortisol changes. The cortisol response examined at 5 and 30 min post-exercise did not appear to be dependent on fence height, while sampling time was shown to significantly affect these changes. Therefore, the sampling times adopted in this study could have influenced the results obtained.

However, the most relevant and significant effect of exercise was recorded during competition in horses when they jumped the highest fences<sup>8</sup>, as also occurred during these experimental performances carried out in this study by the same horses. This again would suggest that, in these experimental trials, the results obtained could be dependent on the sampling times adopted and/or the actual effect of exercise could be also partly masked by the different resting cortisol levels, which were lower before session 2 and session 3 than before session 1. Although the tests were performed using a randomized cross-over design, it is likely that this result is also due to a combined effect of the exercise being performed over consecutive days, inducing stress and fatigue, and a negative feedback effect of increased post-exercise cortisol levels.

As regards the finding that there were no statistically significant effects of exercise on  $\beta$ -endorphin and ACTH concentrations, this could be attributed mainly to the low intensity and speed of the showjumping exercise performed by horses, which was certainly below the critical thresholds for  $\beta$ -endorphin<sup>15</sup> and ACTH<sup>18</sup> secretion. However, this hypothesis cannot be confirmed because of the lack of lactate evaluation. Nevertheless, a peak  $\beta$ -endorphin response was evaluated after warm-up in session 2.

It is known that  $\beta$ -endorphin modifies the excitability of the central nervous system, inducing the control over various functional systems, including motor activity and pain perception following lactic acid and catecholamine increases during exercise<sup>41,42</sup>. Catecholamine and lactate production contribute to HPA axis regulation during exercise and in turn induce stress-coping activities<sup>30</sup>.

In contrast with the results obtained after competitive showjumping over different fence height<sup>8</sup>, which showed that the equivalent increases in the two hypophyseal hormones recorded at 5 min post-experimental exercise were maintained when the fence height increased, in this study the increases in values were higher than after competition for  $\beta$ -endorphin and lower for ACTH.

The lack of significant changes in ACTH and  $\beta$ -endorphin levels after exercise could also be explained by the high individual variability of values shown by horses both before and especially after the exercise, probably explained by the different horses' temperament and consequent individual catecholamine secretion<sup>43</sup>.

Despite this, treadmill exercises at submaximal levels of exercise<sup>13</sup> as well as exercise tests of different intensity<sup>14,15,28,33</sup> were found to induce significant increases in  $\beta$ -endorphin concentrations, which were correlated to exercise intensity, but they too induced a widely varying range of values<sup>14</sup>.

The significant effect of sampling time found for the hormone changes recorded may again explain the lack of significant changes being recorded. In fact, as already stated, in session 2 the time of peak  $\beta$ -endorphin response was evaluated after warm-up, with significantly higher than basal values, although a wide range of individual variability was recorded; and in both session 2 and session 3, the  $\beta$ -endorphin and ACTH concentrations were higher than basal 5 min post-exercise. It is known that the time of peak  $\beta$ -endorphin response varies according to training and horses' age<sup>13,26,27</sup>. However, during incremental exercise tests in Thoroughbreds, plasma ACTH concentrations were increased between 5 and 30 min after the end of exercise<sup>18</sup>, and in previous studies,  $\beta$ -endorphin increases were recorded using a 5-min post-graded exercise test<sup>26</sup> and were statistically significant at 10 min even after submaximal exercise<sup>13</sup>. A peak  $\beta$ -endorphin increase after test at 95%  $VO_{2max}$  was measured in the 10 min after exercise and returned to pre-exercise values 30–90 min after the end of exercise<sup>15</sup>. In contrast, in horses tested at 60%  $VO_{2max}$ ,  $\beta$ -endorphin concentrations returned to pre-exercise values more slowly, 30–120 min after the end of exercise<sup>15</sup>.

In this study, two-way RM-ANOVA demonstrated that, in the experimental conditions adopted, the fence height had no influence on ACTH and  $\beta$ -endorphin changes in the 30 min following the exercise.

The controlled experimental conditions adopted probably also had an influence on the limited hormone changes recorded in this study because, in a previous study with Thoroughbreds exercising on treadmill, the ACTH response was dependent on emotional stress in the form of novel environmental stimuli<sup>31</sup>. In competitive conditions, no clear relationships between  $\beta$ -endorphin changes and fence height could be demonstrated, although the  $\beta$ -endorphin increase was found to be delayed in horses jumping the highest fences with all jumping a penalty round<sup>35</sup>.

The entity of responses of the two hypophyseal hormones to experimental showjumping was not reciprocally correlated.  $\beta$ -Endorphin was correlated to cortisol increase only in session 1, when the exercise presented a low level of difficulty, while ACTH was correlated to cortisol only in session 3, when the horses were subjected to the highest level of exercise stress. In young horses during training, treadmill exercises at submaximal levels of exercise produced increases only in  $\beta$ -endorphin concentrations<sup>13</sup>. In Thoroughbreds exercising on a treadmill, the response of cortisol was not found to be correlated to plasma ACTH<sup>18</sup>.

It was recorded that changes in  $\beta$ -endorphin and cortisol concentrations did not occur in phase when showjumping over the highest fences, in line with results already mentioned in response to competitive exercise<sup>35</sup>, while differing from previous reported data obtained during training<sup>13</sup>. However, a lack of correlation between  $\beta$ -endorphin and cortisol concentrations has also been reported in other situations: in normal and crib-biter horses<sup>44</sup>, after exercise tests associated with thermal stress<sup>45</sup> and after restraint and application of a nasogastric tube<sup>11</sup>, all of which are stressful conditions with a high emotional content. Therefore, it would be interesting to better evaluate whether there is any relationship between  $\beta$ -endorphin changes, emotional content of experimental trials and the effects of sampling times adopted. On the other hand, in humans with obsessive-compulsive disorders<sup>46</sup>, it has been suggested that the elevated cortisol levels may exert a negative feedback effect on  $\beta$ -endorphin release and such a result could explain the hormonal changes recorded after session 2 and session 3.

In conclusion, this study demonstrated that experimental showjumping sessions led to cortisol increases and moderately higher levels than basal of ACTH and  $\beta$ -endorphin after jumping over the highest fences. However, these hormonal modifications were not found to be dependent on fence height based on the sampling times adopted in this study. Hence, these results suggest the need to gather further experimental data in order to differentiate the possible role of the emotional content of stress experienced during exercise from the influence of the sampling times adopted,

to explain hormonal changes and their correlations in the course of showjumping exercises and in order to gain more precise information concerning the correct timing of samples for the post-exercise evaluation of hypophyseal hormones.

## Acknowledgements

Support for this project was provided by the Italian Ministry for Universities and Research (MiUR) and the University of Messina.

## References

- Thornton JR (1985). Hormonal responses to exercise and training. *The Veterinary Clinics of North America: Equine Practice* **1**: 477-495.
- Church DB, Evans DL, Lewis DR and Rose RJ (1987). The effect of exercise on plasma adrenocorticotrophin, cortisol and insulin in the horse and adaptation with training. In: Gillespie JR and Robinson NE (eds) *Equine Exercise Physiology 2*. Davis, CA: ICEEP Publications, pp. 506-515.
- Lekeux P, Art T, Linden A, Desmecht D and Amory H (1991). Heart rate, haematological and serum biochemical responses to show jumping. In: Persson SGB, Lindholm A and Jeffcott LB (eds) *Equine Exercise Physiology 3*. Davis, CA: ICEEP Publications, pp. 385-390.
- Rose RJ and Hodgson DR (1994). Haematology and biochemistry. In: Hodgson DR and Rose RJ (eds) *The Athletic Horse: Principles and Practice of Equine Sports Medicine*. Philadelphia, PA: WB Saunders Comp., pp. 63-78.
- Ferlazzo A and Fazio E (1997). Endocrinological variables in blood and plasma. In: Lindner A (ed.) *Performance Diagnosis of Horses*. Wageningen: Wageningen Academic Publishers, pp. 30-43.
- McKeever KH (2002). Exercise physiology of the older horse. *The Veterinary Clinics of North America: Equine Practice; Geriatrics* **18**: 469-490.
- Hyypä S (2005). Endocrinal responses and regulation in exercising horse. *Livestock Production Science* **92**: 113-121.
- Ferlazzo A, Fazio E and Medica P (2007). Hormonas y Ejercicio. In: Boffi F (ed.) *Fisiologia del Ejercicio en Equinos*. Buenos Aires: Editorial Inter-Médica S.A.I.C.I., pp. 153-164.
- Adler GK (2000). Exercise and fatigue - is neuroendocrinology an important factor? *The Journal of Clinical Endocrinology & Metabolism* **85**: 2167-2169.
- Canali E, Ferrante V, Mattiello S, Sacerdote P, Panerai AE, Lebelt D and Zanella A (1996). Plasma levels of  $\beta$ -endorphin and *in vitro* lymphocyte proliferation as indicators of welfare in horses in normal or restrained conditions. *Pferdebeilkunde* **12**: 415-418.
- Hydbring E, Nyman S and Dahlborn K (1996). Changes in plasma cortisol, plasma  $\beta$ -endorphin, heart rate, haematocrit and plasma protein concentration in horses during restraint and use of a naso-gastric tube. *Pferdebeilkunde* **12**: 423-427.
- McCarthy RN, Jeffcott B and Clarke IJ (1993). Preliminary studies on the use of plasma  $\beta$ -endorphin in horses as an indicator of stress and pain. *Journal of Equine Veterinary Science* **13**: 216-219.
- McCarthy RN, Jeffcott LB, Funder JW, Fullerton M and Clarke IJ (1991). Plasma beta-endorphin and adrenocorticotropin in young horses in training. *Australian Veterinary Journal* **68**: 359-361.
- Mehl ML, Sarkar DK, Schott HC II, Brown JA, Sampson SN and Bayly WM (1999). Equine plasma  $\beta$ -endorphin concentrations are affected by exercise intensity and time of day. *Equine Veterinary Journal Supplement* **30**: 567-569.
- Mehl ML, Schott HC II and Sarkar DK (2000). Effects of exercise intensity and duration on plasma  $\beta$ -endorphin concentrations in horses. *American Journal of Veterinary Research* **61**: 969-973.
- Marc M, Parvizi N, Ellendorff F, Kallweit E and Elsaesser F (2000). Plasma cortisol and ACTH concentrations in the warmblood horse in response to a standardized treadmill exercise test as physiological markers for evaluation of training status. *Journal of Animal Science* **78**: 1936-1946.
- Linden A, Art T, Amory H, Desmecht D and Lekeux P (1991). Effect of 5 different types of exercise, transportation and ACTH administration on plasma cortisol concentration in sport horses. In: Persson SGB, Lindholm A and Jeffcott LB (eds) *Equine Exercise Physiology 3*. Davis, CA: ICEEP Publications, pp. 391-396.
- Nagata S, Takeda F, Kurosawa M, Mima K, Hiraga A, Kai M and Taya K (1999). Plasma adrenocorticotropin, cortisol and catecholamines response to various exercises. *Equine Veterinary Journal Supplement* **30**: 570-574.
- Linden A, Art T, Amory H, Desmecht D and Lekeux P (1990). Comparison of the adrenocortical response to both pharmacological and physiological stresses in sport horses. *Journal of Veterinary Medicine (A)* **37**: 601-604.
- Foreman JH and Ferlazzo A (1996). Physiological responses to stress in the horse. *Pferdebeilkunde* **12**: 401-404.
- Lindner A, Fazio E, Ferlazzo AM, Medica P and Ferlazzo A (2000). Plasma cortisol concentration in Thoroughbred horses during and after standardized exercise tests on a treadmill and effect of conditioning on basal cortisol values. *Pferdebeilkunde* **16**: 502-510.
- Lindner A, Fazio E, Medica P and Ferlazzo A (2002). Effect of age, time record  $V_4$  on plasma cortisol concentration in standardbred racehorses during exercise. *Pferdebeilkunde* **18**: 51-56.
- Golland LC, Evans DL, Stone GM, Tyler CM, Rose RJ and Hodgson DR (1996a). The effect of overtraining on plasma cortisol concentrations at rest and in responses to exercise and administration of synthetic adrenocorticotropin in standardbred racehorses. *Pferdebeilkunde* **12**: 531-533.
- Golland LC, Evans DL, Stone GM, Tyler-McGowan CM, Hodgson DR and Rose RJ (1996b). Plasma cortisol and beta-endorphin concentrations in trained and over-trained standardbred racehorses. *Pflügers Archives of European Journal of Physiology* **439**: 11-17.
- McKenna B, Lambert M and Evans JA (1993). A study of  $\beta$ -endorphin and cortisol levels in the exercising horse. In Foreman, J. (ed.) *Proceedings of the 12th Conference Association for Equine Sports Medicine*. Santa Barbara, CA: Veterinary Practice Publishing Co., pp. 39-43.
- Malinowski K, Shock EJ, Roegner V, Rochelle P, Kearns CF, Guirnalda P and McKeever KH (2002). Age and exercise training alter plasma beta-endorphin, cortisol, and immune parameters in horses. *Journal of Animal Science* **80**(Suppl. 1): 156.
- Malinowski K, Shock EJ, Rochelle P, Kearns CF, Guirnalda PD and McKeever KH (2006). Plasma  $\beta$ -endorphin, cortisol and immune responses to acute exercise are altered by age and exercise training in horses. *Equine Veterinary Journal Supplement* **36**: 267-273.
- Golland LC, Evans DL, Stone GM, Tyler-McGowan CM, Hodgson DR and Rose RJ (1999a). Maximal exercise transiently disrupts hormonal secretory patterns in Standardbred geldings. *Equine Veterinary Journal Supplement* **30**: 581-585.
- Alexander SL, Irvine CH, Ellis MJ and Donald RA (1991). The effect of acute exercise on the secretion of corticotrophin-releasing factor, arginine vasopressin, and

- adrenocorticotropin as measured in pituitary venous blood from the horse. *Endocrinology* **128**: 65-72.
- 30 Kurosawa M, Nagata S, Takeda F, Kurosawa M, Mima K, Hiraga A, Kai M and Taya K (1998). Plasma catecholamine, adrenocorticotropin and cortisol responses to exhaustive incremental treadmill exercise of the Thoroughbred horse. *Journal of Equine Science* **9**: 9-18.
- 31 Hada H, Onaka T, Takahashi T, Hiraga A and Yagi K (2003). Effects of novelty stress on neuroendocrine activities and running performance in thoroughbred horses. *Journal of Neuroendocrinology* **15**: 638-648.
- 32 Goldfarb AH, Hatfield BD, Armstrong D and Potts J (1990). Plasma beta-endorphin concentration: response to intensity and duration of exercise. *Medicine & Science in Sports & Exercise* **22**: 241-244.
- 33 Art T, Franchimont P and Lekeux P (1994). Plasma  $\beta$ -endorphin response in thoroughbred horses to maximal exercise. *The Veterinary Record* **135**: 499-503.
- 34 Clayton HM (1994). Training show jumpers. In: Hodgson DR and Rose RJ (eds) *The Athletic Horse: Principles and Practice of Equine Sports Medicine*. Philadelphia, PA: WB Saunders Comp., pp. 429-438.
- 35 Clayton HM (1989). Terminology for the description of equine jumping kinematics. *Journal of Equine Veterinary Science* **9**: 341-348.
- 36 Ferlazzo A, Fazio E, Aronica V, Di Majo R, Medica P and Grasso L (1998). Circulating concentrations of  $\beta$ -endorphin, ACTH and cortisol in horses after jumping over fences of different size. *Proceedings of the Conference on Equine Sports Medicine and Science Córdoba, Spain*, pp. 53-56.
- 37 Fazio E, Alberghina D, Aronica V, Medica P and Ferlazzo A (2000). Effect of performance success on blood  $\beta$ -endorphin ACTH and cortisol levels in show jumping horses. In: Lindner A (ed.) *The Elite Show Jumper*. Essen: Science Consult, pp. 113-115.
- 38 Pell SM and McGreevy PD (1999). A study of cortisol and beta-endorphin levels in stereotypic and normal Thoroughbreds. *Applied Animal Behaviour Science* **64**: 81-90.
- 39 Cayado P, Muñoz-Escassi B, Domínguez C, Manley W, Olabbarri B, Sánchez de la Muela M, Castejon F, Marañón G and Vara E (2006). Hormone response to training and competition in athletic horses. *Equine Veterinary Journal Supplement* **36**: 274-278.
- 40 McKeever KH and Gordon ME (2004). Endocrine alterations in the equine athlete. In: Hinchcliff KW, Kaneps A and Geor R (eds) *Equine Sports Medicine and Surgery: Basic and Clinical Sciences of the Equine Athlete*. Philadelphia, PA: Elsevier, pp. 793-814.
- 41 Goldfarb AH and Jamurtas AZ (1997). Beta-endorphin response to exercise. *Sports Medicine* **24**: 8-16.
- 42 Munck A, Naray-Fejes-Toth A and Guyre PM (1987). Mechanisms of glucocorticoid actions on the immune system. In: Berczi I and Kovacs K (eds) *Hormone and Immunity*. Lancaster: MTP Press, pp. 20-37.
- 43 Snow DH, Harris RC, MacDonald IA, Forster CD and Marlin DJ (1992). Effects of high-intensity exercise on plasma catecholamines in the thoroughbred horse. *Equine Veterinary Journal* **24**: 462-467.
- 44 McGreevy P and Nicol C (1998). Physiological and behavioural consequences associated with short-term prevention of crib-biting in horses. *Physiology & Behaviour* **65**: 15-23.
- 45 Williams RJ, Marlin DJ, Smith N, Harris RC, Haresign W and Davis Morel MC (2002). Effects of cool and hot humid environmental conditions on neuroendocrine responses of horses to treadmill exercise. *The Veterinary Journal* **164**: 54-63.
- 46 Weizman RM, Gil-Ad I, Hermes H, Munitz H and Laron Z (1990). Immunoreactive  $\beta$ -endorphin, cortisol, and GH plasma levels in obsessive-compulsive disorder. *Clinical Neuropharmacology* **13**: 297-302.

9. Ferlazzo A, Fazio E, Cravana C, Medica P. Changes of circulating total and free iodothyronines in horses after competitive show jumping with different fence height. *J. Equine Vet. Sci* 2014. 34(6):876-881. 10. Veening J.G, Barendregt H.P. The effects of beta-endorphin: State change modification. *Fluids Barriers. CNS* 2015. 34. Ferlazzo A, Medica P, Cravana C, Fazio E. Endocrine changes after experimental showjumping. *Comp. Exerc. Physiol* 2009. 6(2):59-66. Article information Continued. Copyright: © Cravana, et al.

The endocrine and exocrine glands are specialized organs which release small amounts of hormones in our body. These hormones affect many changes in the human body both inside and outside. This article explains the differences between these two very important glands. Contents [hide]. 1 Summary Table. 2 Descriptions. 3 Endocrine vs Exocrine Glands. 4 Video. Summary Table. Endocrine Glands. Exocrine Glands. Release hormones into the bloodstream. To establish how such malfunctions occur, we experimentally induced obstructive jaundice through bile duct ligation (BDL) using rats, measured serum bilirubin, amylase and insulin levels, and examined histological, immunohistochemical and cytological changes in the pancreas at 3 days, 1 week, and 4 weeks after the BDL. Morphometrical analysis was also conducted. Serum amylase levels steeply increased at 3 days, and then decreased at 1 and 4 weeks after the BDL to lower than the control level. In contrast, the number of zymogen granules decreased at 3 days after the BDL, then increased and even